



Prosopis Pod Meal as an Alternative Aquafeed Ingredient

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Abstract

The expanding human population raises the demand for nutritionally-rich food, and fish is the cheapest and best way to meet this requirement. Currently, the stagnation of capture fisheries encourages aquaculture as a means of providing quality proteins to a growing population. A successful aquaculture enterprise incurs a high expenditure of 50-60% on feed costs. Therefore, reducing feed costs by identifying alternative feed ingredients will improve the economic condition of the aquaculture business. Currently, the conventional ingredients used in aquafeed are becoming more expensive, raising the aquafeed's price and thus the overall cost of fish production. Hence, there is a need to transition to low-cost unconventional aquafeed ingredients, and Prosopis pod meal has excellent potential in this context. Prosopis pod meal has a high nutritional value, and it could be substituted conventional ingredients for instance corn meal, wheat flour, rice bran, etc., in aquafeed. This technical review comprises the scope and constraints of employing Prosopis pod meal as an ingredient in aquafeed.

Keywords: Aquafeed, Fish, Plant-based ingredient, Prosopis pod meal, Unconventional ingredient

Introduction

The rapid expansion of aquaculture is giving a strong indication that the aquafeed industry is going to face a crisis in the near future. Fish feed demand is rising along with fish production, and the main challenge facing the aquafeed business is the rising price of conventional feed ingredients (Hardy, 2010). Feed cost accounts around 50-60% of overall operational costs in aquaculture. Hence, reducing the marginal amount of feed cost would increase the profitability of fish farmers (Ebenebe, 2000). The high demand and scarcity of conventional feed ingredients for aquafeed limited their use in aquafeed formulations. The conventional ingredients used in aquafeed raise the feed or production costs. The rising cost of aquafeed can be lowered by using locally available, inexpensive unconventional feed ingredients. Formulating aquafeed using locally available, economically viable and nutritionally acceptable plant ingredients could increase the profit margin of fish farmers (Gatlin III *et al.*, 2007). From this perspective, the Prosopis pod meal has great potential because it is affordable and

nutritionally balanced. The global availability of Prosopis pods is projected to be around 20-40 million metric tonnes (Sawal *et al.*, 2004). The Prosopis tree is relatively easy to cultivate with low water requirement, and yields of Prosopis pods per hectare are higher (2-10 tonnes) (Riveros, 1992; Choge *et al.*, 2007). The abundant availability of the Prosopis pod suggests that it can be utilized as an unconventional ingredient in an aquafeed instead of high-cost conventional ingredients.

Aquafeed Ingredients

Aquaculture intensification is increasing the demand for aquafeed, which accounts for a considerable portion of total operating costs. Aquafeed prices have risen dramatically during the last few decades. The use of costly conventional feed ingredients in the preparation of aquafeed influences the cost of fish production (Gabriel *et al.*, 2007). The rising cost of conventional ingredients drove feed formulators and aquaculturists to explore locally available affordable unconventional ingredients for aquafeed. These ingredients

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are neither prevalent in markets nor regularly used for commercial aquafeed formulations. Moreover, these ingredients are generally not used for human consumption and are byproducts of agricultural and processing industries (Devendra, 1988).

Description and Distribution of Prosopis

Prosopis is a legume belongs to the Fabaceae family, which thrives in arid and semi-arid areas (Shukla and Kasera, 2004). The genus Prosopis comprises 44 species. Some species are important because they grow rapidly, such as *Prosopis juliflora*, *P. africana*, *P. alba*, *P. chilensis*, *P. cineraria*, *P. glandulosa*, *P. hassleri*, *P. nigra* and *P. pallida*. Prosopis trees are found all over the world and are primarily cultivated in Afghanistan, Africa, America, Australia, Brazil, Burma, India, Indonesia, Iran, Oman, Pakistan, Saudi Arabia, Sri Lanka, United Arab Emirates, United States of America, Yemen, etc. (Choge *et al.*, 2007). The tree is known locally as Ghaf (Arabic), Jand (Pakistan) and Jandi or Khejri or Vilayati Babul (India). It is commonly termed the “wonder tree” and “king of the desert” (Jatasra and Paroda, 1981). In India, the prominent species are *P. cineraria* and *P. juliflora*, which are found primarily in Rajasthan, Telangana, Haryana, Punjab, Gujarat, Andhra Pradesh, Karnataka, Maharashtra, Orissa, Uttar Pradesh and Tamil Nadu (Parkash and Hocking, 1985; Kaushik and Kumar, 2003; Khatri *et al.*, 2010). Prosopis pod has excellent nutritive value with high protein, carbohydrate, mineral and vitamin (Malik *et al.*, 2013). Prosopis also has therapeutic utilities such as antimicrobial,

antifungal, antioxidant, anticancer, antihyperlipidemic and antihyperglycemic activities due to the presence of a bioactive component (Pareek *et al.*, 2015). Prosopis pods are primarily utilized as manure and to a lesser extent in animal feed (cattle, camels, sheep, rabbits, goats, poultry and fish) (Riveros, 1992). It is necessary to investigate the influence of Prosopis pod meal-based diets on fish growth and nutrient utilization and standardize their inclusion levels in diets of different fish species without jeopardizing fish growth and health.

Nutritional Composition of Prosopis Pod Meal

The available literature on the nutritional composition of Prosopis pods indicates that it could be a probable energy and protein ingredient for aquafeed (Table 1). Prosopis pods are highly nutritious and palatable (Anttila *et al.*, 1993), and they have been consumed by cattle, sheep, rabbits, goats, sheep, poultry and fish (Silva, 1986; Silva *et al.*, 1990; Pinheira *et al.*, 1993; Ravikala *et al.*, 1993; Mahgoub *et al.*, 2004; Al-Beitawi *et al.*, 2010; Chovatiya *et al.*, 2016; Al-Harathi *et al.*, 2018; Nascimento *et al.*, 2019). Like other plant-based ingredients, the proximate composition of Prosopis pod meal varies with location, soil type, age, season, etc. (Chopra and Hooda, 2001). The variation in the proximate composition of Prosopis pod meal observed in different studies is most likely due to differences in agronomical and environmental situations, diverse species and different analytical methodologies (Girma *et al.*, 2011; Kingori *et al.*, 2011; Tran, 2015; Tran, 2016).

Table 1: Proximate composition (% dry matter basis) of Prosopis pod meal as well as selected conventional ingredients

Variables	Dry matter	Crude protein	Ether extract	Crude fiber	Nitrogen free extract	Total Ash	References
Prosopis pod	91.4	15.2	2.61	18.58	48.97	6.04	Al-Harathi <i>et al.</i> , 2018
Prosopis pod	90.68	12.05	3.2	22.4	48.27	4.76	Chovatiya <i>et al.</i> , 2016
Prosopis pod	90.2	12.8	1.1	18.4	52.8	5.1	Al-Marzooqi <i>et al.</i> , 2015
Prosopis pod	82-94	7.1-18.5	0.4-3.5	12.3-28	46.5-65.3	1.4-6.0	Kingori <i>et al.</i> , 2011
Prosopis pod	89.15	15.43	6.01	14.6	46.98	6.13	Girma <i>et al.</i> , 2011
Prosopis pod	82-92.6	7.1-16.2	0.4-4.0	12.3-27.7	46.3-71.1	1.4-6.0	Choge <i>et al.</i> , 2007
Prosopis pod	91.2	16.5	4.2	16.8	48.3	5.4	Rao and Reddy, 1983
Prosopis pod	89.2	17.77	2.42	19.82	44.44	4.75	Mabrouket <i>et al.</i> , 2008
Prosopis pod	90-93	9.0-17.0	1.2-4.3	16-34	47-61	5-7	Shukla <i>et al.</i> , 1986
Prosopis pod	90.13	13.64	0.72	14.62	53.77	7.38	Yousif, 2012
Corn meal	88	10.2	4.8	2.8	68.6	1.6	NRC, 2011
Rice bran	91	13	7.4	2.3	66.5	1.8	NRC, 2011
Wheat bran	89	14.8	4	9.9	55	5.3	NRC, 2011
Soybean meal	89	44	1.5	7.3	29.9	6.3	NRC, 2011

The crude protein of Prosopis pod meal ranges from 7.1-18.5% on a dry matter basis (Sawal *et al.*, 2004; Kingori *et al.*, 2011). These notable variations in crude protein level could be due to the previously described explanation. The crude protein level of Prosopis pod is comparable to corn meal, rice bran and wheat bran (NRC, 2011). Prosopis pod

contains an equitable amount of essential amino acids (Table 2). *P. juliflora* contains aspartic acid, threonine, cystine, valine, methionine, isoleucine, leucine, tyrosine, phenylalanine, alanine, histidine, lysine, arginine, serine, glutamic acid and glycine amino acids in the concentration of 0.99%, 0.28%, 0.14%, 0.43%, 0.10%, 0.27%, 0.52% 0.29%,

Table 2: Amino acids composition (concentration/ 100 g ingredients) of Prosopis pod meal as well as selected conventional ingredients

Crude protein and amino acids	Prosopis pod ^a	Prosopis pod ^b	Prosopis pod ^c	Corn meal ^d	Rice bran ^d	Wheat bran ^d	Soybean meal ^d
Protein	15.2	20.2	16.5	10.2	13	14.8	44
Arginine	0.67	2.687	0.56	0.4	1	0.64	3.23
Aspartic acid	1.76	NR	0.99	NR	NR	NR	NR
Cysteine	0.17	0.243	0.14	0.19	0.27	0.27	0.7
Glutamic acid	1.28	NR	1.4	NR	NR	NR	NR
Glycine	1.04	0.895	0.51	NR	NR	NR	NR
Histidine	0.2	0.545	0.19	0.25	0.34	0.3	1.17
Isoleucine	0.54	0.545	0.27	0.29	0.44	0.51	1.99
Leucine	1.03	1.333	0.52	1	0.92	0.89	3.42
Lysine	0.46	0.808	0.32	0.26	0.57	0.36	2.83
Methionine	0.12	0.201	0.1	0.18	0.26	0.21	0.61
Phenylalanine	0.4	0.465	0.33	0.42	0.56	0.63	2.18
Proline	1.11	NR	NR	NR	NR	NR	NR
Threonine	0.47	0.485	0.28	0.3	0.48	0.37	1.73
Tryptophan	0.44	0.222	NR	0.07	0.14	0.17	0.61
Tyrosine	0.2	1.172	0.29	NR	0.4	0.43	1.69
Valine	0.67	0.707	0.43	0.42	0.68	0.59	2.4

NR: not reported. (Source: ^aAl-Harathi *et al.*, 2018; ^bHeuze *et al.*, 2015; ^cShukla *et al.*, 1986, ^dNRC, 2011)

0.33%, 0.37%, 0.19%, 0.32%, 0.56%, 0.41%, 1.4% and 0.51%, respectively (Gangal *et al.*, 2009). Al-Harathi *et al.* (2018) also evaluated the essential amino acid composition of *P. juliflora* and detected arginine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, threonine, tryptophan and valine in the percentage of 0.67, 0.20, 0.54, 1.03, 0.46, 0.12, 0.40, 0.47, 0.44 and 0.67, respectively. The differences in Prosopis pod amino acid composition between studies are most likely attributable to differing origins, agricultural and environmental conditions and measurement tools or methodologies (Silvia *et al.*, 2012). Moreover, the amino acid composition of Prosopis pod meal is comparable to those of conventional feedstuffs such as corn meal, rice bran and wheat bran (NRC, 2011).

Prosopis pods are rich in carbohydrates and can be used as a superior energy ingredient in aquafeed. The pod contains approximately 46-65% nitrogen-free extract (NFE) (Escobar *et al.*, 1987), with saccharose (20-25%) and reduced sugar (10-20%) being the major components (Silva, 1986). Prosopis pods have a high proportion of crude fiber levels (14-34%) (Choge *et al.*, 2007; Kingori *et al.*, 2011) and high differences in crude fiber content in pods may be due to variations in age of a tree, a season of pod collection and area of cultivation (Chopra and Hooda, 2001). The NFE level of Prosopis pod meal is compared to corn meal, rice bran, wheat bran and soybean meal, and it observed that the pod meal has a similar NFE content to wheat bran and slightly lesser NFE content than rice bran or corn meal (Table 1).

The ether extract content of pod meal ranges from 0.4 to

6.0%, which is comparable to corn meal and wheat bran (Kingori *et al.*, 2011; Girma *et al.*, 2011). Prosopis pods are concentrated in unsaturated fatty acids (67.21-73.97%), which comprised monounsaturated fatty acids (18.86-37.83%) and polyunsaturated fatty acids (29.37-50.92%). The saturated fatty acid content ranges from 20.14-26.42%, and the saturated fatty acid/ unsaturated fatty acid ratio ranges from 0.27 to 0.40. Palmitic acid (10.8%), oleic acid (35.5%) and linoleic acid (27.9%) are primary saturated, monounsaturated and polyunsaturated fatty acids, respectively (Choge *et al.*, 2007). Prosopis pod meal contains a high amount of essential docosahexaenoic acid (1.17%), which is vital for fish growth, survival and immune development (Table 3) (Al-Harathi *et al.*, 2018).

The total ash level of the pod meal ranges from 3.6 and 6.13%. Prosopis pods are an excellent source of macro- and micro-minerals (Table 4). Calcium, phosphorus, potassium, magnesium and sodium are found in concentrations ranging from 0.26 to 0.66%, 0.13 to 0.20%, 0.73 to 1.69%, 0.34 to 1.20% and 0.01 to 0.05%, respectively. The pod meal contains iron 764 ppm, zinc 69.4 ppm, manganese 33.9 ppm, copper 36.1 ppm, chromium 21.7 ppm, cadmium 7.4 ppm, nickel 9.8 ppm and lead 28.2 ppm. Prosopis pod mineral compositions are consistent with rice bran, wheat bran and soybean meal (NRC, 2011). Talpada *et al.* (1989) revealed that pods collected in winter have superior iron, zinc and copper level than in summer, whereas manganese is not changed by season. Similarly, pods collected from different areas have different mineral content (Chopra and Hooda, 2001).

Table 3: Fatty acid profile (% of total fatty acids) of Prosopis pod meal

Fatty acids	Prosopis pod ^a	Prosopis pod ^b	Prosopis pod ^c
C6:0	0.569	NR	NR
C8:0	3.14	NR	NR
C10:0	0.649	NR	NR
C12:0	1.31	NR	NR
C13:0	2.08	NR	NR
C14:1	1.08	NR	NR
C14:0	0.151	NR	0.16
C15:1	0.944	NR	NR
C15:0	0.777	NR	NR
C16:1	0.308	NR	NR
C16:0	10.8	NR	13.19
C17:0	0.153	NR	NR
C18:3	0.00	NR	NR
C18:2	27.9	NR	50.92
C18:1	35.5	NR	22.74
C18:0	5.39	NR	4.75
C20:2	0.304	NR	NR
C20:0	0.281	NR	NR
C22:0	NR	NR	1.26
C22:1	NR	NR	0.31
C24:0	NR	NR	0.75
DHA C22:6	1.17	NR	NR
THA	92.54	92.45	94.11
SFA	25.55	26.42	20.14
UFA	67.21	66.03	73.97
MUFA	37.83	18.86	23.05
PUFA	29.37	47.17	50.92
SFA/UFA ratio	0.377	0.400	0.272

NR: not reported; DHA: docosahexaenoic acid; TFA: total fatty acids; SFA: saturated fatty acids; UFA: Unsaturated fatty acids; MUFA: monounsaturated fatty acids; PUFA: polyunsaturated fatty acids. (Source: ^aAl-Harhi *et al.*, 2018; ^bChoge *et al.*, 2007; ^cMarangoni and Alli, 1988)

Phytotherapeutic Compounds of Prosopis and Its Uses

Plants and their byproducts have been shown to be classical sources of medicine and act as nutraceuticals when incorporated into aquafeed. Prosopis is well-known as “A Gift of Nature for Pharmacy” because it contains a variety of medicinally valuable bioactive substances or phytoconstituents (Pareek *et al.*, 2015). Various phytoconstituents have been extracted from pods, such as tannins (gallic acid), alkaloids (prosophylline, spicigerine), flavonoids (patuletin, petulitrin, luteolin,

rutin, prasogenin A, B, C, D and E), steroids (campestral, stigmasterol, sitosterol) *etc.* (Bhardwaj *et al.*, 1981; Rani *et al.*, 2013). These components have beneficial effects on animal growth, digestion and metabolism and immunity (Choge *et al.*, 2007). The phytoconstituents of Prosopis act as antimicrobial, antifungal, antioxidant, anthelmintic, anticancer, antihyperlipidemic and antihyperglycemic (Garg and Mittal, 2013; Malik *et al.*, 2013; Pareek *et al.*, 2015).

Antinutritional Factor and Its Amelioration

The application of plant-based ingredients in aquafeed is advantageous by reason of its widespread availability and cheapness. Nevertheless, the occurrence of antinutritional factors and more fiber limited the inclusion of these ingredients in animal feed as this reduces feed intake or nutrient utilization and affects the health status of animals (Francis *et al.*, 2001). Similarly, Prosopis pod meal also contains antinutrient elements, such as saponins, tannins, phenolic compounds, trypsin inhibitors, phytic acid, *etc.*, which need to ameliorate before they can be used in aquafeed (Table 5). Tannins and total phenolic compounds are the principal antinutritional factors present in pods (Saxena and Venkateshwarlu, 1991). Since aquafeed must be made with more than one ingredient, incorporating Prosopis pod meal up to 30% of feed may reduce overall feed tannin levels within permissible limits. Becker and Makkar (1999) state that dietary tannin powder at 0.2% does not alter fish feed intake or growth performance. Similarly, Twibell and Wilson (2004) suggested that feeding pure saponins below 0.26% in aquafeed does not affect fish growth and feed intake. Therefore, incorporating up to 30% Prosopis pod meal into aquafeed has no saponin concerns because the overall feed saponin content reduces below the allowable limit. Prosopis pod meal contains phytic acid; nevertheless, Francis *et al.* (2001) recommended that the allowed level of phytic acid in aquafeed is less than 0.5%, and Prosopis pod meal contains lower than the tolerable limits.

Tannins, phenolic compounds, saponins and phytic acid are soluble in water and can be ameliorated through soaking, cooking, autoclaving and wet heat treatment (Kataria *et al.*, 1989; Vijayakumari *et al.*, 1996). Polyethylene glycol (tannin blocking agent) can also be used in Prosopis pod meal-based aquafeed to mitigate tannin detrimental effects (Makkar and Becker, 1996; Bhatta *et al.*, 2002). Fermentation is another possible method for eliminating or removing tannins, saponins, and phytic acid from the Prosopis pod meal (Mukhopadhyay and Ray, 1999; Bhatt *et al.*, 2011). The supplementation of phytase in fish feed can reduce phytic acid's negative effects (Gabriel *et al.*, 2007). Prosopis pod meal contains trace quantities of trypsin inhibitors, which can be removed through moist thermal processing (Norton, 1991).

Selected Studies on the Application of Prosopis Pods in Fish and Livestock Feed

Several researchers have used Prosopis pods in fish and livestock feed and found that it is a viable alternative to the

Table 4: Minerals composition of Prosopis pod meal as well as selected conventional ingredients

Minerals	Prosopis poda	Prosopis podb	Prosopis podc	Prosopis podd	Prosopis pode	Rice branf	Wheat branf	Soybean mealf
Calcium %	0.66	0.271	0.43	0.26	0.5	0.25	0.13	0.03
Phosphorus %	0.20	NR	0.17	0.13	0.2	0.66	1.16	0.28
Potassium %	NR	1.693	0.73	1.31	0.9	0.28	1.22	0.31
Magnesium (ppm)	0.034	0.119	NR	0.12	0.076	0.20	0.57	0.13
Iron (ppm)	764	289.9	400	NR	99	74.0	145.0	48.0
Zinc (ppm)	69.4	NR	300	NR	1279	68.0	95.0	17.0

NR: not reported. (Source: ^aAl-Harathi *et al.*, 2018; ^bChovatiya *et al.*, 2016; ^cManhique *et al.*, 2017; ^dGirma *et al.*, 2011; ^eKoech *et al.*, 2010; ^fNRC, 2011)

Table 5: Antinutritional factors of Prosopis pod meal

Antinutritional factors	Prosopis pod ^a	Prosopis pod ^b
Total phenol (%)	0.582	0.640
Tannin (%)	0.973	0.860
Phytic acids (%)	0.188	0.181
Saponin (%)	0.393	0.317

(Source: ^aChovatiya *et al.*, 2016; ^bSarasvati *et al.*, 2014)

high-cost limited available conventional feedstuffs (Table 6). Most investigations on amalgamation of Prosopis pod meal as a fish feed ingredient were conducted on tilapia species. Prosopis pod meal can completely replace the conventional high-cost ingredients (corn meal, wheat flour, rice bran and cottonseed meal) in tilapia feed. The available literature suggests that it may include 6.0 to 38.67% of the feed of tilapia feed with positive impacts on growth and health (Mabrouk *et al.*, 2008; Ramos *et al.*, 2012; Sena *et al.*, 2012; Nascimento *et al.*, 2019). Fermented Prosopis pod

Table 6: Selected study on the use of Prosopis pod meal in fish feed

Sl. No.	Species	Replacement	Results	References
1	<i>Oreochromis niloticus</i> juveniles	Corn meal	Corn meal can be replaced by <i>P. juliflora</i> pods meal entirely, without hostile health consequences.	Souza <i>et al.</i> , 2019
2	<i>Oreochromis niloticus</i> juveniles	Corn meal & wheat flour	<i>P. juliflora</i> pod meal could be incorporated in juvenile tilapia diet up to 38.67%, with encouraging effects on growth and health.	Nascimento <i>et al.</i> , 2019
3	<i>Labeo rohita</i> fingerlings	Rice bran	Rice bran can completely replace by fermented <i>P. juliflora</i> pod meal (33.4% of diet), and addition of probiotics significantly improve growth performances.	Chovatiya <i>et al.</i> , 2016
4	<i>Oreochromis niloticus</i> juveniles	Corn meal	<i>P. juliflora</i> pod meal can totally replace corn meal or incorporated 20% of the diet for Nile tilapia.	Silva <i>et al.</i> , 2015
5	<i>Oreochromis niloticus</i> juveniles	Cottonseed & corn meal	Inclusion of <i>P. juliflora</i> pod meal at the level of 15% of feed improves growth performance of tilapia.	Carvalho <i>et al.</i> , 2012
6	<i>Oreochromis niloticus</i> juveniles	Cottonseed meal	<i>P. juliflora</i> pod meal can be incorporated in tilapia diet of up to 20% level without compromising growth performances.	Sena <i>et al.</i> , 2012
7	<i>Valamugil seheli</i> fry	Wheat flour	<i>P. cineraria</i> can be incorporated in Mullet diet up to 20% level without affecting growth performances.	Yousif, 2012
8	<i>Oreochromis niloticus</i> adults	Conventional feedstuffs	The equivalent apparent dry matter and crude protein digestibility coefficients were observed in tilapia fed with 30% cassava leaf meal, cocoa meal and Prosopis pod meal.	Ramos <i>et al.</i> , 2012
9	<i>Colossoma macropomum</i> fingerlings	Corn meal	<i>P. juliflora</i> pod meal could completely replace corn meal in feed without affecting growth performances of Tambaqui, when included in feed at 33.8% levels.	Miranda <i>et al.</i> , 2009
10	<i>Oreochromis niloticus</i> fry	Corn meal	<i>P. juliflora</i> pod meal at 6% dietary level can significantly increase growth compare to non-inclusion groups.	Mabrouk <i>et al.</i> , 2008

can wholly substitute rice bran in the feed of *Labeo rohita* without impacting growth performance and feed intake (Chovatiya *et al.*, 2016). Miranda *et al.* (2009) observed that *P. juliflora* pod meal (33.8%) could be substituted for corn meal in the diet of *Colossoma macroporum* fingerlings. *P. cineraria* can be incorporated up to 20% of the diet for *Valamugil seheli* fry without impairing growth performance (Yousif, 2012).

Prosopis pod meal as a substitute for conventional feedstuffs in terrestrial animals is well established (Table 7) (Sawal *et al.*, 2004). Prosopis pod meal was added to chicken feed to replace maize meal and wheat flour, and shown that it could be blended up to 20% without compromising growth or egg production (Silva, 1986; Al-Beitawi *et al.*, 2010; Girma *et al.*, 2011; Al-Marzooqi *et al.*, 2015; Al-Harhi *et al.*, 2018). Both *P. cinerea* and *P. juliflora* species pods have been added to goat and sheep feed, and it has been observed that the pod meal can substitute conventional feed ingredients (Mahgoub *et al.*, 2004; Koech *et al.*, 2010). Incorporating up to 30% Prosopis pod meal into the diet did not alter the growth performance of lamb and pig finishers (Pinheira *et al.*, 1993; Ravikala *et al.*, 1993).

Fermentation Process Improves Nutritional Quality of Prosopis Pod Meal

Fermentation can improve the nutritional quality of plant-based ingredients. Solid-state fermentation of Prosopis pod meal with *Bacillus subtilis* improves nutritional quality by enhancing essential nutrients and reducing antinutritional factors (Yusuf *et al.*, 2008). The Prosopis pod crude protein can be increased 1.4-fold after 48 hours of fermentation with *B. subtilis*. Prosopis pod meal contains 40 to 52% soluble sugar (sucrose), and fermentation with *B. subtilis* increased the total soluble sugar of pod meal by 28.66% after 24 hours (Cardozo *et al.*, 2010). The pod meal's crude fiber content is as high as 24.64%, which may be lowered to 17.37% after 48 hours by *B. subtilis* fermentation. Prosopis pod meal's antinutritional factors can be decreased or eliminated through fermentation. Phytic acid can be eliminated or reduced to undetectable levels through fermentation, while tannins of pod meal can be reduced by 16% after 96 hours of fermentation with *B. subtilis* (Sarasvati *et al.*, 2014). Chovatiya *et al.* (2016) also found that fermentation of Prosopis pod meal with *Lactobacillus acidophilus* enhanced crude protein levels from 12.05 to 15.08% and eliminated phytic acid entirely.

Table 7: Selected studies on the use of Prosopis pod meal in livestock feed

Sl. No.	Species	Replacement	Results	References
1	Chicken	Corn meal	Prosopis pod meal could be used in the chicken feed as an unconventional feedstuff.	Al-Harhi <i>et al.</i> , 2018
2	Chicken	Maize meal	Dietary Prosopis pod meal at 10% level improves growth and egg production.	Manhique <i>et al.</i> , 2017
3	Chicken	Crushed corn	Prosopis pod meal can be added at level of 5% in chicken feed without affecting growth performance.	Al-Marzooqi <i>et al.</i> , 2015
4	Chicken	Wheat short	In broilers feed ground Prosopis pod meal can be incorporated up to 20% of diet to reduce feed cost.	Girma <i>et al.</i> , 2011
5	Chicken	Maize meal	Prosopis pods meal at 20% levels could be incorporated in diets of chickens with no loss of growth and egg production.	Al-Beitawi <i>et al.</i> , 2010
6	Goat	Conventional feedstuffs	Feeding 200g Prosopis pods per day to individual goat improve weight gain without affecting feed intake and digestibility.	Koech <i>et al.</i> , 2010
7	Sheep and Goat	Conventional feedstuffs	<i>P. cineraria</i> and <i>P. juliflora</i> pod meal can be incorporated up to 15 and 20% in diets of sheep and goats, respectively, without affecting body weight gain and feed conversion ratio.	Mahgoub <i>et al.</i> , 2004
8	Lamb	Wheat bran	The inclusion of less than 30% Prosopis pod meal in diet did not affect growth performance.	Ravikala <i>et al.</i> , 1993
9	Pig finisher	Maize and soybean meal	The pod meal can be included in finisher pigs diet up to 30% level.	Pinheira <i>et al.</i> , 1993
10	Rabbit	Conventional feedstuffs	Nutrient intake and digestibility do not affect by the inclusion of Prosopis pod meal up to 30% of diet.	Silva <i>et al.</i> , 1990
11	Sheep	Cassava	The 100% cassava leaf meal can replace by Prosopis pod meal without effecting feed intake and nutrient digestibility.	Barros <i>et al.</i> , 1986
12	Laying hen	Wheat bran	The wheat bran can completely replace by Prosopis pod meal without affecting feed intake, FCR and egg production.	Silva, 1986

Conclusion

Aquaculture is gaining popularity due to its rapid growth rate among the animal-based protein sector. Nowadays, feed manufacturers are employing a higher proportion of conventional feed ingredients in aquafeed, raising the feed cost. To lower feed costs, shifting from conventional ingredients to readily available and low-cost unconventional ingredients is necessary. Prosopis pod meal can be used as an alternative ingredient in aquafeed. Prosopis pod meal has an excellent nutritional profile and can be used instead of regular ingredients such as corn meal, wheat flour, rice bran, etc. It also contains some antinutritional factors as other plant-based ingredients, although these can be reduced or removed through various processing methods. Fermentation improves the nutritional profile and antinutritional factor of the Prosopis pod meal. Hence, Prosopis pod meal can be incorporated into aquafeed as an alternate feed ingredient to reduce feed costs and reliance on conventional ingredients.

References

- Al-Beitawi, N.A., Awawdeh, F.T., Khwaileh, M.M., 2010. Preliminary study on *Prosopis juliflora* pods as unconventional feed ingredient in diets of broiler chicks. *Animal Nutrition and Feed Technology* 10(1), 51-60.
- Al-Harhi, M.A., Attia, Y.A., Al-Sagan, A.A., Elgandy, M.F., 2018. Nutrients profile, protein quality and energy value of whole prosopis pods meal as a feedstuff for poultry feeding. *Italian Journal of Animal Science* 18(1), 30-38. DOI: <https://doi.org/10.1080/1828051X.2018.1464889>.
- Al-Marzooqi, W., Al-Kharousi, K., Kadim, I.T., Mahgoub, O., Zekri, S., Al-Maqbaly, R., Al-Busaidi, M., 2015. Effects of feeding *Prosopis juliflora* pods with and without exogenous enzyme on performance, meat quality and health of broiler chickens. *International Journal of Poultry Science* 14(2), 76-88. DOI: <https://doi.org/10.3923/ijps.2015.76.88>.
- Anttila, L.S., Alakoski-Johansson, G.M., Johansson, S.G., 1993. Browse preference of Orma livestock and chemical composition of *Prosopis juliflora* and nine indigenous woody species in Bura, Eastern Kenya. *East African Agricultural and Forestry Journal* 58, 83-90.
- Barros, N.A.M., Bai, O.A., Fonesca, F.C.E., 1986. Use of *Prosopis juliflora* (SW) DC and cassava (*Manihot utilissima* Phol) for confined sheep feeding during dry season. In: *II International Conference on The current state of knowledge on Prosopis juliflora*. Recife, Brazil. August 25-29, 1986. pp. 379-383. URL: <https://www.fao.org/3/ad317e/AD317E14.htm#ch6.6>.
- Becker, K., Makkar, H.P.S., 1999. Effects of dietary tannic acid and quebracho tannin on growth performance and metabolic rates of common carp (*Cyprinus carpio* L.). *Aquaculture* 175(3-4), 327-335. DOI: [https://doi.org/10.1016/S0044-8486\(99\)00106-4](https://doi.org/10.1016/S0044-8486(99)00106-4).
- Bhardwaj, D.K., Gupta, A.K., Jain, R.K., Sharma, G.C., 1981. Chemical examination of *Prosopis spicigera* seeds. *Journal of Natural Products* 44(6), 656-659. DOI: <https://doi.org/10.1021/np50018a003>.
- Bhatt, S.S., Chovatiya, S.G., Shah, A.R., 2011. Evaluation of raw and hydrothermally processed *Prosopis juliflora* seed meal as supplementary feed for the growth of *Labeo rohita* fingerlings. *Aquaculture Nutrition* 17(2), 164-173. DOI: <https://doi.org/10.1111/j.1365-2095.2009.00745.x>.
- Bhatta, R., Shinde, A.K., Vaithiyanathan, S., Sankhyan, S.K., Verma, D.L., 2002. Effect of polyethylene glycol-6000 on nutrient intake, digestion and growth of kids browsing *Prosopis cineraria*. *Animal Feed Science and Technology* 101(1-4), 45-54. DOI: [https://doi.org/10.1016/S0377-8401\(02\)00180-3](https://doi.org/10.1016/S0377-8401(02)00180-3).
- Cardozo, M.L., Ordoñez, R.M., Zampini, I.C., Cuello, A.S., Dibenedetto, G., Isla, M.I., 2010. Evaluation of antioxidant capacity, genotoxicity and polyphenol content of non-conventional foods: Prosopis flour. *Food Research International* 43(5), 1505-1510. DOI: <https://doi.org/10.1016/j.foodres.2010.04.004>.
- Carvalho, J.S.O., Azevedo, R.V.D., Ramos, A.P.D.S., Braga, L.G.T., 2012. Agroindustrial byproducts in diets for Nile tilapia juveniles. *Revista Brasileira de Zootecnia* 41(3), 479-484. DOI: <https://doi.org/10.1590/S1516-35982012000300002>.
- Choge, S.K., Pasiecznik, N.M., Harvey, M., Wright, J., Awan, S.Z., Harris, P.J.C., 2007. Prosopis pods as human food, with special reference to Kenya. *Water SA* 33(3), 419-424. URL: <https://hdl.handle.net/10520/EJC116431>.
- Chopra, D., Hooda, M.S., 2001. Variability in chemical composition of *Prosopis juliflora* seeds and hull. *Indian Journal of Animal Sciences* 18(3), 282-284.
- Chovatiya, S., Bhatt, S., Shah, A., Dube, P., 2016. An investigation on the use of *Prosopis juliflora* pods as a carbohydrate source supplemented with probiotics in the diet of *Labeo rohita* fingerlings. *Iranian Journal of Fisheries Sciences* 17(2), 327-345.
- Devendra, C., 1988. General approaches to animal nutrition research and their relevance to fish production in the Asian region. In: *Finfish Nutrition Research in Asia*. (Ed.) de Silva, S.S. Heinemann Asia, Singapore. pp. 7-24.
- Ebenebe, C.I., 2000. Mini livestock production in Nigeria, The present and future. In: *Proceedings of 5th Annual Conference of Animal Science Association of Nigeria-2000*. Port Harcourt, Nigeria, September 13-15, 2000. pp. 19-22.
- Escobar, B., Rumeo, M., Boeza, G., Sato, X., Vasqufz, M., 1987. Characterization and chemical composition of mesquite (*Prosopis chilensis*) pods. *Revista Chilena de Nutrición* 15(2), 113-116.
- Francis, G., Makkar, H.P., Becker, K., 2001. Antinutritional factors present in plant-derived alternate fish feed ingredients and their effects in fish. *Aquaculture* 199(3-4), 197-227. DOI: [https://doi.org/10.1016/S0044-8486\(01\)00526-9](https://doi.org/10.1016/S0044-8486(01)00526-9).
- Gabriel, U.U., Akinrotimi, O.A., Bekibele, D.O., Onunkwo, D.N., Anyanwu, P.E., 2007. Locally produced fish feed:

- potentials for aquaculture development in subsaharan Africa. *African Journal of Agricultural Research* 2(7), 287-295.
- Gangal, S., Sharma, S., Rauf, A., 2009. Fatty acid composition of *Prosopis cineraria* seeds. *Chemistry of Natural Compounds* 45(5), 705-707. DOI: <https://doi.org/10.1007/s10600-009-9425-8>.
- Garg, A., Mittal, S.K., 2013. Review on *Prosopis cineraria*: A potential herb of Thar desert. *Drug Invention Today* 5(1), 60-65. DOI: <https://doi.org/10.1016/j.dit.2013.03.002>.
- Gatlin III, D.M., Barrows, F.T., Brown, P., Dabrowski, K., Gaylord, T.G., Hardy, R.W., Herman, E., Hu, G., Krogdahl, Å., Nelson, R., Overturf, K., 2007. Expanding the utilization of sustainable plant products in aquafeeds: a review. *Aquaculture Research* 38(6), 551-579. DOI: <https://doi.org/10.1111/j.1365-2109.2007.01704.x>.
- Girma, M., Urge, M., Animut, G., 2011. Ground *Prosopis juliflora* pods as feed ingredient in poultry diet: effects on growth and carcass characteristics of broilers. *International Journal of Poultry Science* 10(12), 970-976. DOI: <https://doi.org/10.3923/ijps.2011.970.976>.
- Hardy, R.W., 2010. Utilization of plant proteins in fish diets: effects of global demand and supplies of fishmeal. *Aquaculture Research* 41(5), 770-776. DOI: <https://doi.org/10.1111/j.1365-2109.2009.02349.x>.
- Heuze, V., Tran, G., Boval, M., Renaudeau, D., 2015. Mesquite (*Prosopis juliflora*). Feedipedia, a programme by INRA, CIRAD, AFZ and FAO. Available at: <http://feedipedia.org/node/554>. Accessed on: 25th October, 2019.
- Jatasra, D.S., Paroda, R.S., 1981. *Prosopis cineraria*--an unexploited treasure of the Thar Desert. *Forage Research* 7(1), 1-12.
- Kataria, A., Chauhan, B.M., Punia, D., 1989. Antinutrients and protein digestibility (in vitro) of mungbean as affected by domestic processing and cooking. *Food Chemistry* 32(1), 9-17. DOI: [https://doi.org/10.1016/0308-8146\(89\)90003-4](https://doi.org/10.1016/0308-8146(89)90003-4).
- Kaushik, N., Kumar, V., 2003. Khejri (*Prosopis cineraria*)-based agroforestry system for arid Haryana, India. *Journal of Arid Environments* 55(3), 433-440. DOI: [https://doi.org/10.1016/S0140-1963\(02\)00289-6](https://doi.org/10.1016/S0140-1963(02)00289-6).
- Khatri, A., Rathore, A., Patil, U.K., 2010. *Prosopis cineraria* (L.) druce: a boon plant of desert-an overview. *International Journal of Biomedical and Advance Research* 1(5), 141-149.
- Kingori, A.M., Odero-Waitituh, J.A., Guliye, A.Y., 2011. Mathenge (*Prosopis juliflora*): an underutilized livestock feed resource in Kenya. *Research Journal of Animal Sciences* 5(4-6), 43-51. DOI: <https://doi.org/10.3923/rjnasci.2011.43.51>.
- Koech, O.K., Kinuthia, R.N., Wahome, R.G., Choge, S.K., 2010. Effects of *Prosopis juliflora* seed pod meal supplement on weight gain of weaner Galla goats in Kenya. *Research Journal of Animal Sciences* 4(2), 58-62. DOI: <https://doi.org/10.3923/rjnasci.2010.58.62>.
- Mabrouk, H., Hilmi, E., Abdullah, M., 2008. Nutritional value of *Prosopis juliflora* pods in feeding Nile tilapia (*Oreochromis niloticus*) fries. *Arab Gulf Journal of Scientific Research* 26(1-2), 49-62.
- Mahgoub, O., Kadim, I.T., Al Ajmi, D.S., Al Saqry, N.M., Al Abri, A.S., Richie, A.R., Al Halhali, A.S., Forsberg, N.E., 2004. Use of local range tree (*Prosopis* spp.) pods in feeding sheep and goats in the Sultanate of Oman. In: *Nutrition and Feeding Strategies of Sheep and Goats under Harsh Climates*. CIHEAM, (Options Méditerranéennes: Série A. Séminaires Méditerranéens), Hammamet, Tunisia. pp. 191-195.
- Makkar, H.P.S., Becker, K., 1996. Effect of pH, temperature, and time on inactivation of tannins and possible implications in detannification studies. *Journal of Agricultural and Food Chemistry* 44(5), 1291-1295. DOI: <https://doi.org/10.1021/jf9506287>.
- Malik, S., Mann, S., Gupta, D., Gupta, R.K., 2013. Nutraceutical properties of *Prosopis cineraria* (L.) Druce pods: A component of Panchkuta. *Journal of Pharmacognosy and Phytochemistry* 2(2), 66-73.
- Manhique, A.J., King'ori, A.M., Wachira, A.M., 2017. Effect of ground mature *Prosopis juliflora* pods inclusion in layer diets on performance of improved indigenous chicken in Kenya. *Livestock Research for Rural Development* 29(19), 266-272.
- Marangoni, A., Alli, I., 1988. Composition and properties of seeds and pods of the tree legume *Prosopis juliflora* (DC). *Journal of the Science of Food and Agriculture* 44(2), 99-110. DOI: <https://doi.org/10.1002/jsfa.2740440202>.
- Miranda, E.C. de, Guimarães, I.G., Cabral Junior, C.R., Pinheiro, D.M., 2009. Growth performance of tambaqui (*Colossoma macropomum*) fed diets containing different replacement levels of corn by mesquite pods meal. *Pubvet* 3(2), 25-34.
- Mukhopadhyay, N., Ray, A.K., 1999. Effect of fermentation on the nutritive value of sesame seed meal in the diets for rohu, *Labeo rohita* (Hamilton), fingerlings. *Aquaculture Nutrition* 5(1), 229-236. DOI: <https://doi.org/10.1046/j.1365-2095.1999.00101.x>.
- Nascimento, A.A., Melo, J.F.B., de Souza, A.M., Melo, F.V.S.T., 2019. Inclusion of mesquite pod meal (*Prosopis juliflora*) in diets for Nile tilapia (*Oreochromis niloticus*) juveniles. *Boletim do Instituto de Pesca* 45(3), 1-9. DOI: <https://doi.org/10.20950/1678-2305.2019.45.3.425>.
- Norton, G., 1991. Proteinase inhibitors. Chapter 4. In: *Toxic Substances in Crop Plants*. (Eds.) D'Mello, J.P.F., Duffus, C.M. and Duffus, J.H. Woodhead Publishing, Sawston, Cambridge. pp. 68-106. DOI: <https://doi.org/10.1533/9781845698454.68>.
- NRC (National Research Council), 2011. *Nutrient Requirements of Fish and Shrimp*. 7th Rev. Ed. The National Academies Press, Washington DC. pp. 70-76. DOI: <https://doi.org/10.17226/13039>.
- Pareek, A.K., Garg, S., Kumar, M., 2015. *Prosopis cineraria*: a gift of nature for pharmacy. *International Journal of Pharmaceutical Sciences and Research* 6(6), 958-964.
- Parkash, R., Hocking, D., 1985. Some favourite trees for fuel

- and fodder. In: *Society for Promoting of Wastelands Development*, New Delhi, India. p. 187. URL: <https://www.cabi.org/isc/abstract/19870617791>.
- Pinheira, M.J.P., de Sousa, R.P., Espindole, G.B., Desousa, R.P., 1993. Effect of adding mesquite pods to diets of finishing swine. *Pesquisa Agropecuaria Brasileira* 28(12), 1443-1449.
- Ramos, A.P.S., Braga, L.G.T., Carvalho, J.S.O., de Oliveira, S.J.R., 2012. Digestibility of agro-industrial byproducts in 200 and 300-g Nile tilapia. *Revista Brasileira de Zootecnia* 41(2), 462-466. DOI: <https://doi.org/10.1590/S1516-35982012000200032>.
- Rani, B., Singh, U., Sharma, R., Gupta, A., Dhawan, N.G., Sharma, A.K., Sharma, S., Maheshwari, R.K., 2013. *Prosopis cineraria* (L) Druce: A desert tree to brace livelihood in Rajasthan. *Asian Journal of Pharmaceutical Research and Health Care* 5(2), 58-64.
- Rao, N.S.R., Reddy, M.R., 1983. Utilization of *Prosopis juliflora* pods in the concentrate feeds of cattle and sheep. *Indian Journal of Animal Research* 53(4), 367-372.
- Ravikala, K., Patel, A.M., Murthy, K.S., Desai, M.C., 1993. Rumen metabolites in growing lambs on feeding *Prosopis juliflora* based complete feeds. *Indian Journal of Animal Sciences* 10(3), 177-180.
- Riveros, F., 1992. The genus *Prosopis* and its potential to improve livestock production in arid and semi arid regions. *Legume Trees and Other Fodder Trees as Protein Sources for Livestock*. FAO, Rome, pp. 257-276.
- Sarasvati, S., Sujata, B., Amita, S., 2014. Effects of fermentation on nutritional quality of *Prosopis juliflora* pods as alternative fish feed. *Research Journal of Animal, Veterinary and Fishery Sciences* 2(12), 1-7.
- Sawal, R.K., Ratan, R., Yadav, S.B.S., 2004. Mesquite (*Prosopis juliflora*) pods as a feed resource for livestock-A review. *Asian-Australasian Journal of Animal Sciences* 17(5), 719-725. DOI: <https://doi.org/10.5713/ajas.2004.719>.
- Saxena, S.K., Venkateshwarlu, J., 1991. Mesquite: an ideal tree for desert reclamation and fuel wood production. *Indian Farming* 41(7), 15-21.
- Sena, M.F., de Azevedo, R.V., Ramos, A.P.S., Carvalho, J.S.O., Costa, L.B., Braga, L.G.T., 2012. Mesquite bean and cassava leaf in diets for Nile tilapia in growth. *Acta Scientiarum: Animal Sciences* 34(3), 231-237. DOI: <https://doi.org/10.4025/actascianimsci.v34i3.13175>.
- Shukla, J.K., Kaseria, P.K. 2004. Response of irrigation and spacing levels on growth parameters of Khejri (*Prosopis cineraria*). *Indian Journal of Forestry* 27(4), 335-338.
- Shukla, P.C., Talpada, P.M., Pande, M.B., Desai, M.C., Desai, H.B., 1986. *Prosopis juliflora* pods and their utilization as cattle feed. In: *Proceedings of Seminar on Prosopis species in Wasteland Development*. Surendrabag, Gujarat, India, March 24-26, 1986. pp. 143-146.
- Silva, T.R.M., Chung, S., de Araújo, T.A.T., de Azevedo, K.S.P., dos Santos, M.C., Bicudo, Á.J.A., 2015. Replacing corn by mesquite meal (*Prosopis juliflora*) in diets for juvenile Nile tilapia reared in low temperature. *Revista Brasileira de Ciências Agrárias* 10(3), 460-465. DOI: <https://doi.org/10.5039/agraria.v10i3a4168>.
- Silva, A.D.A., Rodrigues, M.E., de Silva, J.F., 1990. Nutritional value of mesquite beans (*Prosopis juliflora*) in the diet of rabbits. *Veterinária Zootecnia* 2, 9-16.
- Silva, S., 1986. *Prosopis juliflora* (SW) DC in Brazil. In: *International Conference on Prosopis*. Recife, Brazil, September 14-17, 1986. pp. 25-29.
- Silvia, D., Masturah, M.F., Tajul Aris, Y., Wan Nadia, W.A., Rajeev, B., 2012. The effects of different extraction temperatures of the screw press on proximate compositions, amino acid contents and mineral contents of *Nigella sativa* meal. *American Journal of Food Technology* 7(4), 180-191. DOI: <https://doi.org/10.3923/ajft.2012.180.191>.
- Souza, A.M., Silva, A.T., Felix e Silva, A., Campeche, D.F.B., Melo, J.F.B., Vidal, L.V.O., 2019. Mesquite bean (*Prosopis juliflora*) meal in diets of Nile tilapia (*Oreochromis niloticus*): Nutritional value, growth, physiological responses and health. *Aquaculture Research* 50(1), 49-62. DOI: <https://doi.org/10.1111/are.13867>.
- Talpada, P.M., Desai, H.B., Patel, Z.N., Shukla, P.C., 1989. Seasonal variation in trace element content of pods of *Prosopis juliflora*. *Agricultural Science Digest* 9(2), 68-70.
- Tran, G., 2015. Syrian mesquite (*Prosopis farcta*). Feedipedia, a programme by INRA, CIRAD, AFZ and FAO. Available at: <http://feedipedia.org/node/262>. Accessed on: 14th October, 2019.
- Tran, G., 2016. Chilean mesquite (*Prosopis chilensis*). Feedipedia, a programme by INRA, CIRAD, AFZ and FAO. Available at: <http://feedipedia.org/node/553>. Accessed on: 12th December, 2019.
- Twibell, R.G., Wilson, R.P., 2004. Preliminary evidence that cholesterol improves growth and feed intake of soybean meal-based diets in aquaria studies with juvenile channel catfish, *Ictalurus punctatus*. *Aquaculture* 236(1-4), 539-546. DOI: <https://doi.org/10.1016/j.aquaculture.2003.10.028>.
- Vijayakumari, K., Siddhuraju, P., Janardhanan, K., 1996. Effect of domestic processing on the levels of certain antinutrients in *Prosopis chilensis* (Molina) Stunz. seeds. *Food Chemistry* 59(3), 367-371. DOI: [https://doi.org/10.1016/S0308-8146\(96\)00249-X](https://doi.org/10.1016/S0308-8146(96)00249-X).
- Yousif, O.M., 2012. The potential of dehydrated enteromorpha, *Prosopis cineraria* Linn. (Leguminosae) pods meal and date pits in formulated feed for bluespot grey mullet fry, *Valamugil seheli* (Forsskal). *Journal of Agriculture and Veterinary Science* 13(1), 36-45.
- Yusuf, N.D., Ogah, D.M., Hassan, D.I., Musa, M.M., Doma, U.D., 2008. Effect of decorticated fermented prosopis seed meal (*Prosopis africana*) on growth performance of broiler chicken. *International Journal of Poultry Science* 7(11), 1054-1057. DOI: <https://doi.org/10.3923/ijps.2008.1054.1057>.